

The Effects of Alpha Interferon on the Development of Autoimmune Thyroiditis in the NOD H2h4 Mouse

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Alpha interferon (αIFN) therapy is known to induce thyroid autoimmunity in up to 40% of patients. The mechanism is unknown, but Th1 switching has been hypothesized. The aim of our study was to examine whether aIFN accelerated the development of thyroiditis in genetically susceptible mice. We took advantage of NOD-H2h4, a genetically susceptible animal model, which develops thyroiditis when fed a high iodine diet. Six to eight week old male NOD H2h4 mice were injected with mouse αIFN (200 units) or with saline three times a week for 8 weeks. All mice drank iodinated water (0.15%). Mice were sacrificed after 8 weeks of injection. Their thyroids were examined for histology and blood was tested for antithyroglobulin antibody levels. T4 and glucose levels were also assessed. In the IFN-injected group, 6/13 (46.2%) developed thyroiditis and/or thyroid antibodies while in the saline-injected group, only 4/13 (30.8%) developed thyroiditis and/or thyroid antibodies (p = 0.4). The grade of thyroiditis was not different amongst the two groups. None of the mice developed clinical thyroiditis or diabetes mellitus. Our results showed that α IFN treatment did not accelerate thyroiditis in this mouse model. This may imply that α IFN induces thyroiditis in a non-genetically dependent manner, and this would not be detected in a genetically susceptible mouse model if the effect were small. Alternatively, it is possible that α IFN did not induce thyroiditis in mice because, unlike in humans, in mice α IFN does not induce Th1 switching.

Keywords: Alpha interferon; Autoimmunity; Animal model; Thyroiditis

INTRODUCTION

Alpha interferon (αIFN) treatment for various medical conditions is known to induce thyroid autoimmunity in up to 40% of patients. The association between αIFN treatment and autoimmune thyroiditis was reported as early as 1986 by Burman et al. (1986), who studied patients receiving aIFN for carcinoid tumors. The association is most recognized in patients with hepatitis C (Watanabe et al., 1994), although it has been noted to occur in several other conditions treated by aIFN including breast cancer, carcinoid and hematologic malignancies (Fentiman et al., 1988; Ronnblom et al., 1991a,b; Gisslinger et al., 1992; Silvestri et al., 1994; Watanabe et al., 1994). The thyroid diseases associated with α IFN therapy include both autoimmune hyperthyroidism and hypothyroidism (Koh et al., 1997). In up to 40% of cases, thyroid dysfunction persists after discontinuation of the α IFN therapy (Koh *et al.*, 1997).

Factors that predispose patients to the development of α IFN-induced thyroiditis include hepatitis C infection, positive pre-treatment antithyroid antibodies, dose and duration of α IFN therapy, genetic factors, stress, dietary iodine intake, female gender, pregnancy and HLA-A2 in the Japanese population (Preziati *et al.*, 1995; Roti *et al.*, 1996; Custro *et al.*, 1997; Koh *et al.*, 1997; Minelli *et al.*, 1997; Fernandez-Soto *et al.*, 1998; Dumoulin *et al.*, 1999; Kakizaki *et al.*, 1999, 2000). The mechanisms by which α IFN induces thyroid autoimmunity are unclear.

We hypothesized that α IFN accelerates subclinical thyroiditis by immune modulation in genetically predisposed individuals. In order to examine this hypothesis, we took advantage of the NOD H2h4 mouse model, a model in which the mice are genetically prone to develop

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thyroiditis when placed on iodinated drinking water (Rasooly *et al.*, 1996; Braley-Mullen *et al.*, 1999). We used the NOD H2h4 mouse model to examine the effects of α IFN on animals genetically susceptible to thyroiditis.

MATERIALS AND METHODS

Animal Protocol

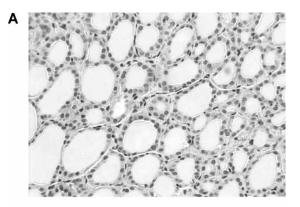
Mouse interferon alpha was obtained from PBL Biomedical Laboratories (New Brunswick, NJ). Six to eight week old NOD H2h4 male mice were purchased from Taconic Farms (Germantown, NY). Male mice were used in our experiments since previous studies (Rasooly et al., 1996; Braley-Mullen et al., 1999) and our own experiments revealed that male mice are more prone to thyroiditis in this model (data not shown). Thirteen mice were injected intra peritoneally (i.p.) with aIFN (200 units, 0.4 cc) three times a week for eight weeks. As controls, 13 mice were injected with saline (0.4 cc) three times a week for eight weeks. After eight weeks of injections, mice were sacrificed, 1 ml of blood was drawn, and their thyroids were removed. All these 26 mice were given 0.15% iodinated drinking water [Sodium Iodide, Sigma Chemical Company (St. Louis, MO)]. Additional 14 mice (6 injected with saline and 8 injected with α IFN) received regular water without iodine supplementation.

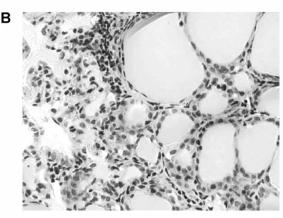
Measurement of Murine Tg (mTg) Antibodies

Serum was analyzed for mTg antibodies by an ELISA assay as previously described (Imaizumi et al., 2001). Briefly, 96 well-plates (Immulon 1, Dynex Technologies Inc., Pittsburgh, PA) were coated with mouse thyroglobulin (0.1 mcg thyroglobulin/well diluted in Carbonate/Bicarbonate Buffer, pH 9.6) and incubated overnight at 4°C. The plates were washed with 0.05% PBS-Tween and nonspecific binding was blocked for 30 min with 3% BSA-PBS/Tween. After washing, the sera were diluted 1:100 in 0.5% BSA/PBS and added to the plates for 2h at room temperature. Plates were washed again 6 times and alkaline phosphatase conjugated anti mouse IgG (Sigma Diagnostics, St. Louis, MO) was added for 30 min. Plates were washed 4 times and then developed using p-nitrophenyl phosphate as substrate. Absorbance was read at 405 nm. Samples were considered positive if the OD was 2 standard deviation (SD) above the average OD for controls.

Thyroid Histology

Mice thyroids were fixed in 10% formalin and stained by hematoxylin and eosin. The severity of thyroiditis was graded on a scale from +1 to +4.0 as follows: +1, small focal areas of lymphocytic cells; +2, focal collection of lymphocytic cells with some follicular destruction; +3, diffuse lymphocytic infiltration of thyroid follicles





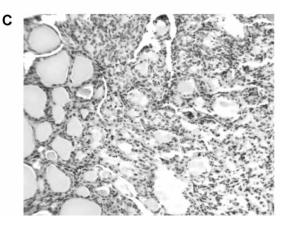


FIGURE 1 Histological grading of thyroiditis. After eight weeks injections, mice were sacrificed. Thyroids were removed and fixed and stained by H&E. (A) Normal mouse thyroid, (B) Grade 1 thyroiditis, (C) Grade 2 thyroiditis. None of our mice developed thyroiditis with severity > +2.

involving approximately 40% or less of the thyroid and +4, >40% lymphocytic infiltration of thyroid follicles (Fig. 1; Imaizumi *et al.*, 2001).

Measurement of Total Thyroxine (T4) and Glucose Levels

T4 was measured by radioimmunoassay using the neonatal T4 Coat-A-Count kit (Diagnostics Products Corporation, Los Angeles, CA). Glucose levels were measured using the One Touch Profile Glucometer (Lifescan Inc., Milpitas, CA).

Data Analysis

Data were analyzed using Student's, Chi-Square and Fisher Exact tests. Probability values less than 0.05 were considered significant.

RESULTS

Effects of Iodine on Development of Thyroiditis

None of the 14 mice that did not receive iodine supplementation (6 injected with saline and 8 injected with α IFN) developed thyroiditis. In contrast, 10/26 (38.5%) of the mice that received iodine supplementation developed thyroiditis (p = 0.007).

Autoantibody Levels in α IFN-injected Mice and Controls

Positive thyroglobulin antibodies were detected in 3/13 (23%) of the mice injected with α IFN compared to 0% of the control mice injected with saline (p = 0.07).

Thyroid Histology in aIFN-injected Mice and Controls

Thyroid infiltration was detected in 5/13 (38.5%) of the α IFN-injected mice and 4/13 (30.8%) in the control mice (Table I). The grade of thyroiditis was not different between the α IFN-injected versus controls. When we used positive thyroid antibodies and/or positive histology to define thyroiditis, 6/13 (46.2%) of the mice injected with α IFN developed thyroiditis compared to 4/13 (30.8%) in the control mice (Table I). This difference did not reach statistical significance (p=0.4).

Clinical Disease in aIFN-injected Mice and Controls

None of the mice developed evidence of clinical thyroid dysfunction and/or diabetes mellitus. There were no significant differences in the T4 levels between the 4 groups mice (Fig. 2). Moreover, the levels of T4 were similar in the mice that developed thyroiditis and those that did not (average Total T4 level = 4.24 in mice that developed thyroiditis vs. 4.19 in mice that did not develop thyroiditis) (Fig. 2).

Since it has been previously reported that insulitis develops in 20-30% of NODH2h4 mice

TABLE I Thyroiditis and thyroglobulin antibody frequency in mice injected with saline and αIFN

	Saline injection (n=13)	αIFN injection (<i>n</i> =13)	P value
Positive histology for thyroiditis	4 (31%)	5 (39%)	NS*
Thyroiditis on histology and/or elevated Tg-Ab's	4 (31%)	6 (46%)	NS

^{*} NS, not significant.

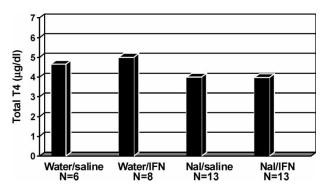


FIGURE 2 T4 levels in mice injected with IFN or saline and fed with water or NaI. There were no significant differences in the T4 levels of mice that developed thyroiditis and those that did not in both groups (average Total T4 level = 4.24 vs. 4.19).

(Braley-Mullen *et al.*, 1999), we also tested blood glucose levels. There were no significant differences in the glucose levels between mice that developed thyroiditis and those that did not in both groups (average glucose level = 174 vs. 165.2) (Fig. 3).

DISCUSSION

Interferons are cytokines that are involved in immune modulation, possessing both antiviral and antitumoral activity. Interferons are grouped into Types I and II. Type I interferons are acid stable and include alpha, beta and omega subtypes. Type II interferons are acid labile and include the gamma subtype (Walter *et al.*, 1998). α IFN first became available for use in humans in the form of leukocyte-derived interferon in the mid 1970's. Subsequently, in 1986, recombinant α IFN became available for clinical use. Since that time, recombinant interferons have been used to treat a wide spectrum of medical conditions, including hepatitis, and several cancers (Walter *et al.*, 1998).

Interferons were shown to induce and/or exacerbate several autoimmune diseases, including autoimmune thyroiditis, autoimmune hepatitis, rheumatoid and lupus like diseases, and autoimmune diabetes mellitus. Interestingly, thyroid disease is the most prevalent autoimmune disease exacerbated or induced by α IFN (Dumoulin *et al.*, 1999). It is unclear whether this reflects the higher prevalence of thyroid autoimmunity in the general population or a thyroid specific effect of interferon.

Our hypothesis was that αIFN accelerates thyroiditis in genetically prone individuals. Evidence supporting our hypothesis is that most of the risk factors for αIFNinduced thyroiditis are genetic (e.g. positive pre-treatment antithyroid antibodies, and female gender) (Preziati *et al.*, 1995; Roti *et al.*, 1996; Custro *et al.*, 1997; Minelli *et al.*, 1997; Tunbridge *et al.*, 1977; Koh *et al.*, 1999; Fernandez-Soto *et al.*, 1998; Kakizaki *et al.*, 1999; Dumoulin *et al.*, 1999; Kakizaki *et al.*, 2000). In order to

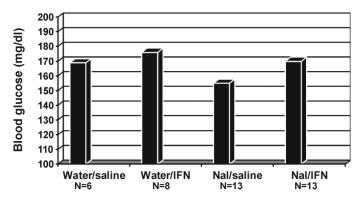


FIGURE 3 Glucose levels in mice injected with IFN or saline and fed with water or NaI. There were no significant differences in the glucose levels between mice that developed thyroiditis and those that did not in both groups (average glucose level = 174 vs. 165.2).

test our hypothesis, we gave αIFN to mice genetically susceptible to thyroiditis.

Our study revealed that aIFN did not accelerate thyroiditis in this thyroiditis-prone mouse model. Thus our study does not support an accelerating effect of aIFN on autoimmune thyroid disease development at least in our mouse model. Therefore, it is possible that α IFN induces thyroiditis de novo and does not accelerate disease development in genetically predisposed individuals. Such a de novo effect would not be detected in our mice, which are already prone for thyroiditis, especially if this is a mild effect. Alternatively, it is possible that α IFN did accelerate thyroiditis development in the NOD H2h4 mice but that the effect was too small to be detected in our experiments. Our studies showed a higher frequency of thyroiditis in the α IFN injected group versus the controls (48.2% and 30.8%, respectively), however this difference did not reach statistical significance. Power calculations show that at least 80 mice per group would have been required to reach statistical significance for the difference that we observed. Additionally, there is some variability in the incidence of thyroiditis in this mouse model, which may have interfered with detection of a small effect (Rasooly et al., 1996; Braley-Mullen et al., 1999). Another explanation for the lack of an effect of aIFN in the mice may be due to its inability to induce Th1 switching in mice. Unlike in humans, aIFN does not induce Th1 switching in mice because of its failure to activate the signal transducer and activator of transcription 4 (Stat4) (Farrar and Murphy, 2000). Some authors have suggested that αIFN accelerates human thyroiditis by inducing Th1 switching (Farrar and Murphy, 2000) and, if this is indeed the case, the lack of αIFN-induced thyroiditis in our mice may reflect the absence of aIFN-induced Th1 differentiation and activation of Stat4 in mice. However, it is unclear whether Th1 switching can explain αIFN-induced thyroiditis as 40% of patients develop Graves' disease, generally believed to be a Th2 mediated disease (Davies, 2000).

In summary, α IFN did not accelerate thyroiditis in this mouse model. However, minor α IFN effects cannot be excluded. Additional studies are needed to examine this possibility.

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